

Restriction Endonucleases

Bioron GmbH offers 74 restriction endonucleases purified by SibEnzyme Ltd. (www.sibenzyme.com). All enzymes are available in bulk and retail quantities. If you need other enzymes produced by SibEnzyme we are pleased to help you. Please contact us for details.

| | 5'→3' | | 5'→3' | | 5'→3' |
|----------|---------------|----------|-----------|-----------------------|-----------------|
| Aau I | T↓GTACA | BstDS I | C↓CRYGG | Psp124B I | GAGCT↓C |
| Acc16 I | TGC↓GCA | BstFN I | CG↓CG | PspE I | G↓GTNACC |
| Acc65 I | G↓GTACC | BstH2 I | RGCGC↓Y | Pst I | CTGCA↓G |
| Acc113 I | AGT↓ACT | BstHH I | GCG↓C | Pvu II | CAG↓CTG |
| AccBS I | GAG↓CGG | BstHP I | GTT↓AAC | Rsa I | GT↓AC |
| Acs I | R↓AATTY | BsuR I | GG↓CC | Sal I | G↓TCGAC |
| Afe I | AGC↓GCT | CciN I | GC↓GGCCGC | Sbf I | CCTGCA↓GG |
| Ahl I | A↓CTAGT | Dra I | TTT↓AAA | Sfi I | GGCCN>NNN↓NGGCC |
| Alu I | AG↓CT | EcoR I | G↓AATTC | Sfr274 I | C↓TCGAG |
| Apa I | GGGCC↓C | EcoR V | GAT↓ATC | Sfr303 I | CCGC↓GG |
| AspS9 I | G↓GNCC | FauND I | CA↓TATG | Sma I | CCC↓GGG |
| AsuNH I | G↓CTAGC | Hind III | A↓AGCTT | Smi I | ATTT↓AAAT |
| BamH I | G↓GATCC | Hinf I | G↓ANTC | Sph I | GCATG↓C |
| Bgl I | GCCN>NNN↓NGGC | Hpa II | C↓CGG | Sse9 I | ↓AATT |
| Bgl II | A↓GATCT | HspA I | G↓CGC | Ssp I | AAT↓ATT |
| Bme18 I | G↓GWCC | Kpn I | GGTAC↓C | Taq I | T↓CGA |
| Bpu14 I | TT↓CGAA | Ksp22 I | T↓GATCA | Tru9 I | T↓TAA |
| Bsa29 I | AT↓CGAT | Kzo9 I | ↓GATC | Vha464 I | C↓TTAAG |
| Bsc4 I | CCN>NNN↓NNGG | Mlu I | A↓CGCGT | Vne I | G↓TGCAC |
| Bse8 I | GATNN↓NNATC | MroN I | G↓CCGGC | Vsp I | AT↓TAAT |
| Bse21 I | CC↓TNAGG | Msp I | C↓CGG | Xba I | T↓CTAGA |
| Bse118 I | R↓CCGGY | Nru I | TCG↓CGA | Zsp2 I | ATGCA↓T |
| Bsp19 I | C↓CATGG | Pce I | AGG↓CCT | | |
| Bss NA I | GTA↓TAC | Pci I | A↓CATGT | | |
| BssT1 I | C↓CWWGG | Ple19 I | CGAT↓CG | Site-specific Nickase | |
| Bst4C I | ACN↓GT | PspOM I | G↓GGCCC | N.Bst9 | GAGTCN>NNN↓NN |



Restriction Endonucleases

Genomic Tools

Bioron GmbH offers you several "rare-cutters" - restriction endonucleases especially suitable for genomic research. All these enzymes passed through additional tests for successful and specific cleavage of high molecular weight DNA.

| Sfi I | Cat# | Pack Size |
|----------------|--------|-----------|
| GGCCNNNN↓NGGCC | 244005 | 500 U |
| CCGGN↑NNNNCCGG | 244025 | 2500 U |

| Smi I | Cat# | Pack Size |
|-----------|--------|-----------|
| ATTT↓AAAT | 247010 | 1000 U |
| TAAA↑TTTA | 247050 | 5000 U |

| Sbf I | Cat# | Pack Size |
|-----------|--------|-----------|
| CCTGCA↓GG | 243002 | 200 U |
| GG↑ACGTCC | 243010 | 1000 U |

| CciN I (iso- NotI) | Cat# | Pack Size |
|--------------------|--------|-----------|
| GC↓GGCCGC | 220002 | 200 U |
| CGCCGG↑CG | 220010 | 1000 U |

Site-specific Nickase N.BstT9

A unique tool for generation of DNA fragments with long sticky ends.

Description:

As an additional tool for genomic DNA fragmentation Bioron offers a unique instrument - site-specific Nickase N.Bst9. In contrast to the restriction enzymes, this enzyme cleaves only one DNA strand after the recognition sequence. Meanwhile, if two recognition sequences appear in close vicinity to each other, the enzyme acts exactly as a restriction endonuclease with 10bp-long recognition sequence.

Purified from *Bacillus stearothermophilus*.

Concentration: 2000-10000 units/ml.

Site-specific Nickase N.Bst9

GAGTCNNNN↓NN.....NNNNNGACTC
CTCAGNNNNNN.....NN↑NNNCTGAG

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of T7 DNA with 8 units of the enzyme at 55°C.

| Site-specific nickase N.Bst9 | Cat# | Pack Size |
|------------------------------|--------|-----------|
| GAGTCNNNN↓NN | 257001 | 100 U |
| CTCAGNNNNNN | 257005 | 500 U |



Alphabetic List of Recognition Sequences

All recognition sequences are written 5' to 3' with the point of cleavage indicated by ↓

| | | | | | |
|--------------|----------|----------------|----------------|----------|----------|
| A↓AGCTT | Hind III | CC↓TNAGG | Bse21 I | GGGCC↓C | Apa I |
| AAT↓ATT | Ssp I | C↓CWWGG | BssT1 I | G↓GNCC | AspS9 I |
| ↓AATT | Sse9 I | CGAT↓CG | Ple19 I | G↓GTACC | Acc65 I |
| A↓CATGT | Pci I | CG↓CG | BstFN I | GGTAC↓C | Kpn I |
| A↓CGCGT | Mlu I | C↓TCGAG | Sfr274 I | G↓GTNACC | PspE I |
| ACN↓GT | Bst4C I | CTGCA↓G | Pst I | G↓GWCC | Bme18 I |
| A↓CTAGT | Ahl I | C↓TTAAG | Vha464 I | GT↓AC | Rsa I |
| A↓GATCT | Bgl II | G↓AATTC | EcoR I | GTA↓TAC | BssNA I |
| AGC↓GCT | Afe I | GAG↓CGG | AccBS I | G↓TCGAC | Sal I |
| AG↓CT | Alu I | GAGCT↓C | Psp124B I | G↓TGCAC | Vne I |
| AGG↓CCT | Pce I | GAGTCNNNN↓NN | Nickase Bst9 I | GTT↓AAC | BstHP I |
| AGT↓ACT | Acc113 I | G↓ANTC | Hinf I | R↓AATTY | Acs I |
| AT↓CGAT | Bsa29 I | GAT↓ATC | EcoR V | R↓CCGGY | Bse118 I |
| ATGCA↓T | Zsp2 I | ↓GATC | Kzo9 I | RGC GC↓Y | BstH2 I |
| AT↓TAAT | Vsp I | GATNN↓NNATC | Bse8 I | T↓CGA | Taq I |
| ATTT↓AAAT | Smi I | GCATG↓C | Sph I | TCG↓CGA | Nru I |
| CAG↓CTG | Pvu II | G↓CCGGC | MroN I | T↓CTAGA | Xba I |
| CA↓TATG | FauND I | GCCNNNN↓NGGC | Bgl I | T↓GATCA | Ksp22 I |
| C↓CATGG | Bsp19 I | G↓CGC | HspA I | TGC↓GCA | Acc16 I |
| CCC↓GGG | Sma I | GCG↓C | BstHH I | T↓GTACA | Aau I |
| CCGC↓GG | Sfr303 I | GC↓GGCCGC | CciN I | T↓TAA | Tru9 I |
| C↓CGG | Hpa II | G↓CTAGC | AsuNH I | TTA↓TAA | Psi I |
| C↓CGG | Msp I | G↓GATCC | BamH I | TT↓CGAA | Bpu14 I |
| CCNNNNN↓NNGG | Bsc4 I | GG↓CC | BsuR I | TTT↓AAA | Dra I |
| C↓CRYGG | BstDS I | GGCCNNNN↓NGGCC | Sfi I | | |
| CCTGCA↓GG | Sbf I | G↓GGCCC | PspOM I | | |

R= A or G
 Y= T or C
 W= A or T
 N = A or C or G or T

Restriction Endonucleases

List of Prototype enzymes and Isoschizomers from Bioron

| Well-known isoschizomers | Recognition sequence | Restriction endonuclease from Bioron | Well-known isoschizomers | Recognition sequence | Restriction endonuclease from Bioron |
|--------------------------|----------------------|--------------------------------------|--------------------------|----------------------|--------------------------------------|
| ApaI I | G↓TGCAC | Vne I | Nco I | C↓CATGG | Bsp19 I |
| Apo I | R↓AATTY | Acs I | Nde I | CA↓TATG | FauND I |
| Ase I | AT↓TAAT | Vsp I | Nhe I | G↓CTAGC | AsuNH I |
| Ava II | G↓GWCC | Bme18 I | Not I | GC↓GGCCGC | CciN I |
| Bcl I | T↓GATCA | Ksp22 I | Nsi I | ATGCA↓T | Zsp2 I |
| Bsl I | CCNNNNN↓NNGG | Bsc4 I | Pae I | GCATG↓C | Sph I |
| BspLU11 I | A↓CATGT | Pci I | Pvu I | CGAT↓CG | Ple19 I |
| Bst4C I | ACN↓GT | Bst4C I | Sac I | GAGCT↓C | Psp124B I |
| BstE II | G↓GTNACC | PspE I | Sac II | CCGC↓GG | Sfr303 I |
| Cfr10 I | R↓CCGGY | Bse118 I | Sau3A I, Mbo I | ↓GATC | Kzo9 I |
| Cla I | AT↓CGAT | Bsa29 I | Sau96 I | G↓GNCC | AspS9 I |
| Dsa I | C↓CRYGG | BstDS I | Sca I | AGT↓ACT | Acc 113 I |
| Eco47 III | AGC↓GCT | Afe I | Spe I | A↓CTAGT | Ahl I |
| Fsp I | TGC↓GCA | Acc 16 I | Sse8387 I | CCTGCA↓GG | Sbf I |
| Hae II | RGCGC↓Y | BstH2 I | Stu I | AGG↓CCT | Pce I |
| Hae III | GG↓CC | BsuR I | Sty I | C↓CWWGG | BssT1 I |
| Hha I | GCG↓C | BstHH I | Swa I | ATTT↓AAAT | Smi I |
| Hin6 I | G↓CGC | HspA I | Tsp509 I | ↓AATT | Sse9 I |
| Hpa I | GTT↓AAC | BstHP I | Xho I | C↓TCGAG | Sfr274 I |
| Mse I | T↓TAA | Tru9 I | | | |

Unit Definition:

One unit of restriction endonuclease activity is defined as the amount of enzyme required to completely digest 1 µg of substrate DNA in a total reaction volume of 50 µl in one hour using the buffer provided. Incubations are performed at the appropriate incubation temperature as indicated on the Technical Data Sheet.

Quality Control:

The results of all quality control assays are reported on the Technical Data Sheet provided with each enzyme.

Ligation of DNA fragments:

DNA fragments are produced by the excessive over-digestion of substrate DNA with each restriction

endonuclease. These fragments are then ligated with T4 DNA ligase at a 5' termini concentration of 0.1-1.0 µM. The ligated fragments are then recut with the same restriction endonuclease. A normal banding pattern after cleavage indicates that both 3' and 5' termini are intact and the enzyme preparation is free of detectable exonucleases and phosphatases.

Overnight assay for nonspecific nuclease contamination:

All restriction endonucleases are incubated overnight in the recommended buffer with 1 µg of substrate DNA in a volume of 50 µl. The characteristic banding pattern produced by the enzyme in one hour is compared to the pattern produced by the excess of enzyme incubated overnight.

All enzymes are supplied with an appropriate reaction buffer. For the majority of restriction enzymes we recommend to use 5 main buffers (B, G, O, W, Y), special buffers are used for EcoR I and Ksp22 I. The composition of the buffers can be found in the table below. Please find additional information in the buffer activity chart on page 43.

| Buffers | Composition |
|---------|---|
| B | 10mM TrisHCl, pH 7.6, 10mM MgCl ₂ , 1mM DTT |
| G | 10mM TrisHCl, pH 7.6, 10mM MgCl ₂ , 50mM NaCl, 1mM DTT |
| O | 50mM TrisHCl, pH 7.6, 10mM MgCl ₂ , 100mM NaCl, 1mM DTT |
| W | 10mM TrisHCl, pH 8.5, 10mM MgCl ₂ , 100mM NaCl, 1mM DTT |
| Y | 33mM Tris-acetate, pH 7.9, 10mM Mg-acetate, 66mM KCl, 1mM DTT |
| 2W | 20mM TrisHCl, pH 8.5, 10mM MgCl ₂ , 200mM NaCl, 1mM DTT |
| Eco RI | 100mM TrisHCl, pH 7.6, 10mM MgCl ₂ , 50mM NaCl, 1mM DTT |
| 2K | 10mM TrisHCl, pH 7.6, 10mM MgCl ₂ , 200mM KCl, 1mM DTT |
| N. Bst9 | 10mM TrisHCl, pH 8.5, 10mM MgCl ₂ , 150mM KCl, 1mM DTT, 0.1mg/ml BSA |

Restriction Endonucleases

Aau I

| Aau I | Cat# | Pack size |
|---------|--------|-----------|
| T↓GTACA | 258005 | 500 U |
| ACATG↑T | 258025 | 2500 U |

Purified from *Arthrobacter aureescens*.

Concentration: 15000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 15 units of enzyme at 37°C.



Acc16 I

| Acc16 I | Cat# | Pack Size |
|---------|--------|-----------|
| TGC↓GCA | 284002 | 200 U |
| ACG↓CGT | 284010 | 1000 U |

Purified from *Acinetobacter calcoaceticus* 16.

Isoschizomers: Fsp I, Nsb I.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 10 units of enzyme at 37°C.

Note! Activity may be blocked by overlapping dem-methylation.



Restriction Endonucleases

| Acc65 I | Cat# | Pack Size |
|---------|--------|-----------|
| G↓GTACC | 205010 | 1000 U |
| CCATG↑G | 205050 | 5000 U |

Acc65 I

Purified from *Acinetobacter calcoaceticus*.

Concentration: 30-50000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 30 units of enzyme at 37°C.

Note! Activity may be blocked by overlapping dcm methylation.



| Acc113 I | Cat# | Pack Size |
|----------|--------|-----------|
| AGT↓ACT | 259006 | 600 U |
| TCA↑TGA | 259030 | 3000 U |

Acc113 I

Purified from *Acinetobacter calcoaceticus* 113.

Isoschizomer: Sca I.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 70% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 10 units of enzyme at 37°C.



| AccBS I | Cat# | Pack Size |
|---------|--------|-----------|
| GAG↓CGG | 283010 | 1000 U |
| CTC↑GCC | 283050 | 5000 U |

AccBS I

Purified from *Acinetobacter calcoaceticus* BS.

Concentration: 5000-20000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 10 units of enzyme at 37°C.



| Acs I | Cat# | Pack Size |
|---------|--------|-----------|
| R↓AATTY | 206005 | 500 U |
| YTTAA↑R | 206025 | 2500 U |

Acs I

Purified from *Arthrobacter citreus*.

Concentration: 20000-50000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 50 units of enzyme at 37°C.



| | | | | | | | | | | | |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|
| 37°C | Optimal reaction temperature | λ | Substrate DNA for activity test | W | Optimal reaction buffer | Yes | Inactivation at 65°C, 20 min | No | No Inactivation at 65°C, 20 min | 80°C | Inactivation at 80°C, 20 min |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|

Afe I

| Afe I | Cat# | Pack Size |
|---------|--------|-----------|
| AGC↓GCT | 207002 | 200 U |
| TCG↑CGA | 207010 | 1000 U |

Purified from *Alcaligenes faecalis* T2774.

Isoschizomer: Eco47 III.

Concentration: 10000-50000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 80% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of

1 µg of λ DNA with 10 units of enzyme at 37°C.



Ahl I

| Ahl I | Cat# | Pack Size |
|---------|--------|-----------|
| A↓CTAGT | 262010 | 1000 U |
| TGATC↓A | 262050 | 5000 U |

Purified from *Alteromonas haloplanktis* SP.

Isoschizomer: Spe I.

Concentration: 30000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of T7 DNA with 100 units of enzyme at 37°C.



Alu I

| Alu I | Cat# | Pack Size |
|-------|--------|-----------|
| AG↓CT | 208004 | 400 U |
| TC↑GA | 208020 | 2000 U |

Purified from *Arthrobacter luteus*.

Concentration: 1000-3000 units/ml.

Ligation/recutting assay:

After 2-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 4 units of enzyme at 37°C.



Apa I

| Apa I | Cat# | Pack Size |
|---------|--------|-----------|
| GGGCC↓C | 209010 | 1000 U |
| C↑CCGGG | 209050 | 5000 U |

Purified from *Acetobacter pasteurianus*.

Concentration: 10000-30000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 40 units of enzyme at 37°C.

Note! Cleavage by ApaI may be blocked by overlapping dcm-methylation.



Restriction Endonucleases

| AspS9 I | Cat# | Pack Size |
|---------|--------|-----------|
| G↓GNCC | 263010 | 1000 U |
| CCNG↑G | 263050 | 5000 U |

AspS9 I

Purified from *Arthrobacter species S9*.

Isoschizomer: Sau96 I.

Concentration: 40000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 40 units of enzyme at 37°C.



| AsuNH I | Cat# | Pack Size |
|---------|--------|-----------|
| G↓CTAGC | 211005 | 500 U |
| CGATC↑G | 211025 | 2500 U |

AsuNH I

Purified from *Actinobacillus suis NH*.

Isoschizomer: Nhe I.

Concentration: 5000-20000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 10 units of the enzyme at 37°C.



| BamH I | Cat# | Pack Size |
|---------|--------|-----------|
| G↓GATCC | 212040 | 4000 U |
| CCTAG↑G | 212200 | 20000 U |

BamH I

Purified from *E.coli* strain that carries *BamHI* gene from *Bacillus amiloliquefaciens*.

Concentration: 50000 - 150000 units/ml.

Ligation/recutting assay:

After 50-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16 h incubation of 1 µg of λ DNA with 200 units of enzyme at 37°C.



| Bgl I | Cat# | Pack Size |
|--------------|--------|-----------|
| GCCNNNN↓NGGC | 265010 | 1000 U |
| CGGN↑NNNNCCG | 265050 | 5000 U |

Bgl I

Purified from *Bacillus globigii*.

Concentration: 5000-30000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of enzyme at 37°C.



| | | | | | | | | | | | |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|
| 37°C | Optimal reaction temperature | λ | Substrate DNA for activity test | W | Optimal reaction buffer | Yes | Inactivation at 65°C, 20 min | No | No Inactivation at 65°C, 20 min | 80°C | Inactivation at 80°C, 20 min |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|

Bgl II

| Bgl II | Cat# | Pack Size |
|---------|--------|-----------|
| A↓GATCT | 213010 | 1000 U |
| TCTAG↑A | 213050 | 5000 U |

Purified from *Bacillus globigii*.

Concentration: 10000-50000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 30 units of enzyme at 37°C.



Bme18 I

| Bme18 I | Cat# | Pack Size |
|---------|--------|-----------|
| G↓GWCC | 266010 | 1000 U |
| CCWG↑G | 260050 | 5000 U |

Purified from *Bacillus megaterium 18*.

Isoschizomer: Ava II.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of enzyme at 37°C.



Bpu14 I

| Bpu14 I | Cat# | Pack Size |
|---------|--------|-----------|
| TT↓CGAA | 267010 | 1000 U |
| AAGC↓TT | 267050 | 5000 U |

Purified from *Bacillus pumilis 14*.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of enzyme at 37°C.



Bsa29 I

| Bsa29 I | Cat# | Pack Size |
|---------|--------|-----------|
| AT↓CGAT | 214010 | 500 U |
| TAGC↑TA | 214050 | 2500 U |

Purified from *Bacillus stearothermophilus 29*.

Isoschizomer: Cla I.

Concentration: 20000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 30 units of enzyme at 37°C.

Note! Cleavage by Bsa29 I may be blocked by overlapping dam-methylation.



Restriction Endonucleases

| Bsc4 I | Cat# | Pack Size |
|--------------|--------|-----------|
| CCNNNNN↓NNGG | 268005 | 500 U |
| GGNN↑NNNNNCC | 268025 | 2500 U |

Bsc4 I

Purified from *Bacillus schlegelii* 4.

Isoschizomer: Bsl I.

Concentration: 20000-60000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 60 units of enzyme at 55°C.



| Bse21 I | Cat# | Pack Size |
|----------|--------|-----------|
| CC↓TNAGG | 269005 | 500 U |
| GGANT↑CC | 269025 | 2500 U |

Bse21 I

Purified from *Bacillus species* 21.

Concentration: 10000-30000 units/ml.

Ligation/recutting assay:

After 2-fold overdigestion with the enzyme, more than 50% of the DNA fragments may be ligated and more than 95% recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 30 units of enzyme at 37°C.



| Bse8 I | Cat# | Pack Size |
|-------------|--------|-----------|
| GATNN↓NNATC | 270010 | 1000 U |
| CTANN↑NNTAG | 270050 | 5000 U |

Bse8 I

Purified from *Bacillus species* 8.

Concentration: 10000-30000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 80% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 30 units of enzyme at 60°C.



| Bse118 I | Cat# | Pack Size |
|----------|--------|-----------|
| R↓CCGGY | 271002 | 200 U |
| YGGCC↑R | 271010 | 1000 U |

Bse118 I

Purified from *Bacillus stearothermophilus* 118.

Isoschizomer: Cfr10 I.

Concentration: 2000 units/ml.

Ligation/recutting assay:

After 2-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 2 units of enzyme at 65°C.



| | | | | | | | | | | | |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|
| 37°C | Optimal reaction temperature | λ | Substrate DNA for activity test | O | Optimal reaction buffer | Yes | Inactivation at 65°C, 20 min | No | No Inactivation at 65°C, 20 min | 80°C | Inactivation at 80°C, 20 min |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|



Bsp19 I

| Bsp19 I | Cat# | Pack Size |
|---------|--------|-----------|
| C↓CATGG | 215005 | 500 U |
| GGTAC↑C | 215025 | 2500 U |

Purified from *Bacillus species 19*.

Isoschizomer: Nco I.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.



Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 5 units of the enzyme at 37°C.

BssNA I

| BssNA I | Cat# | Pack Size |
|---------|--------|-----------|
| GTA↓TAC | 272010 | 1000 U |
| CAT↑ATG | 272050 | 5000 U |

Purified from *Bacillus stearothermophilus NA*.

Concentration: 30000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.



Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 30 units of enzyme at 37°C.

BssT1 I

| BssT1 I | Cat# | Pack Size |
|---------|--------|-----------|
| C↓CWWGG | 273010 | 1000 U |
| GGWWC↑C | 273050 | 5000 U |

Purified from *Bacillus stearothermophilus T1*.

Isoschizomer: Sty I.

Concentration: 20000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.



Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of enzyme at 60°C.

Note! The excess of the enzyme may cause star activity.

Bst4C I

| Bst4C I | Cat# | Pack Size |
|---------|--------|-----------|
| ACN↓GT | 216002 | 200 U |
| TG↑NCA | 216010 | 1000 U |

Purified from *Bacillus stearothermophilus 4C*.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, less than 5% of the DNA fragments may be ligated.



Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of the enzyme at 65°C.

Restriction Endonucleases

| BstDS I | Cat# | Pack Size |
|---------|--------|-----------|
| C↓CRYGG | 274010 | 1000 U |
| GGYRC↓C | 274050 | 5000 U |

BstDS I

Purified from *Bacillus stearothermophilus* DS.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of enzyme at 65°C.



| BstFN I | Cat# | Pack Size |
|---------|--------|-----------|
| CG↓CG | 275003 | 300 U |
| GC↑GC | 275015 | 1500 U |

BstFN I

Purified from *Bacillus stearothermophilus* FN.

Concentration: 2000-10000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 95% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of enzyme at 60°C.



| BstH2 I | Cat# | Pack Size |
|---------|--------|-----------|
| RGCGC↓Y | 217005 | 500 U |
| Y↑CGCGR | 217025 | 2500 U |

BstH2 I

Purified from *Bacillus stearothermophilus* H2.

Isoschizomer: Hae II.

Concentration: 10000-30000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 40 units of the enzyme at 65°C.



| BstHH I | Cat# | Pack Size |
|---------|--------|-----------|
| GCG↓C | 218020 | 2000 U |
| C↑GCG | 218100 | 10000 U |

BstHH I

Purified from *Bacillus stearothermophilus* HH.

Isoschizomer: Hha I.

Concentration: 50000 units/ml.

Ligation/recutting assay:

After 40-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 100 units of the enzyme at 50°C.



| | | | | | | | | | | |
|------|------------------------------|---|---------------------------------|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|
| 37°C | Optimal reaction temperature | λ | Substrate DNA for activity test | Optimal reaction buffer | Yes | Inactivation at 65°C, 20 min | No | No Inactivation at 65°C, 20 min | 80°C | Inactivation at 80°C, 20 min |
|------|------------------------------|---|---------------------------------|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|

BstHP I

| BstHP I | Cat# | Pack Size |
|---------|--------|-----------|
| GTT↓AAC | 277005 | 500 U |
| CAA↑TTG | 277025 | 2500 U |

Purified from *Bacillus stearothermophilus* HP.

Isoschizomer: Hpa I.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 60% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 30 units of enzyme at 37°C.



BsuR I

| BsuR I | Cat# | Pack Size |
|--------|--------|-----------|
| GG↓CC | 219010 | 1000 U |
| CC↑GG | 219050 | 5000 U |

Purified from *Bacillus subtilis* R.

Isoschizomer: Hae III.

Concentration: 50000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 100 units of the enzyme at 37°C.



CciN I

| CciN I | Cat# | Pack Size |
|-----------|--------|-----------|
| GC↓GGCCGC | 220002 | 200 U |
| CGCCGG↑CG | 220010 | 1000 U |

Purified from *Curtobacterium citreus* N.

Isoschizomer: Not I.

Concentration: 5000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of Adenovirus-2 DNA with 20 units of the enzyme at 37°C.



Dra I

| Dra I | Cat# | Pack Size |
|---------|--------|-----------|
| TTT↓AAA | 221005 | 1000 U |
| AAA↑TTT | 221025 | 5000 U |

Purified from *Deinococcus radiophilus*.

Concentration: 10000-30000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 70% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of the enzyme at 37°C.



| | | | | | | |
|------------------------------|---------------------------------|-------------------------|-----|------------------------------|---------------------------------|------------------------------|
| Optimal reaction temperature | Substrate DNA for activity test | Optimal reaction buffer | Yes | Inactivation at 65°C, 20 min | No Inactivation at 65°C, 20 min | Inactivation at 80°C, 20 min |
|------------------------------|---------------------------------|-------------------------|-----|------------------------------|---------------------------------|------------------------------|

Restriction Endonucleases

| EcoR I | Cat# | Pack Size |
|---------|--------|-----------|
| G↓AATTC | 222050 | 5000 U |
| CTTAA↑G | 222250 | 25000 U |

Purified from E.coli strain that carries *EcoRI* gene from *Escherichia coli*.

Concentration: 20000-50000 units/ml.



EcoR I

Ligation/recutting assay:

After 40-fold overdigestion with EcoRI, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 100 units of enzyme at 37°C.

| EcoR V | Cat# | Pack Size |
|---------|--------|-----------|
| GAT↓ATC | 223010 | 1000 U |
| CTA↑TAG | 223050 | 5000 U |

Purified from *Escherichia coli*.

Concentration: 40000 units/ml.



EcoR V

Ligation/recutting assay:

After 20-fold overdigestion with EcoRV, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 40 units of the enzyme at 37°C.

| FauND I | Cat# | Pack Size |
|---------|--------|-----------|
| CA↓TATG | 278005 | 500 U |
| GTAT↑AC | 278025 | 2500 U |

Purified from *Flavobacterium aquatili ND*.

Isoschizomer: Nde I.

Concentration: 20000 units/ml.



FauND I

Ligation/recutting assay:

After 3-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 6 units of enzyme at 37°C.

Note! The enzyme is very sensitive to DNA impurities.

| Hind III | Cat# | Pack Size |
|----------|--------|-----------|
| A↓AGCTT | 224020 | 5000 U |
| TTCGA↑A | 224100 | 25000 U |

Purified from *Haemophilus influenzae Rd*.

Concentration: 20000-50000 units/ml.



Hind III

Ligation/recutting assay:

After 30-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 60 units of enzyme at 37°C.

| | | | | | | | | | | | |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|
| 37°C | Optimal reaction temperature | λ | Substrate DNA for activity test | W | Optimal reaction buffer | Yes | Inactivation at 65°C, 20 min | No | No Inactivation at 65°C, 20 min | 80°C | Inactivation at 80°C, 20 min |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|



Hinf I

| Hinf I | Cat# | Pack Size |
|--------|--------|-----------|
| G↓ANTC | 225010 | 1000 U |
| CTNA↑C | 225050 | 5000 U |

Purified from *Haemophilus influenzae*.

Concentration: 10000-50000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 80 units of the enzyme at 37°C.



Hpa II

| Hpa II | Cat# | Pack Size |
|--------|--------|-----------|
| C↓CGG | 226005 | 500 U |
| GGC↑C | 226025 | 2500 U |

Purified from *Haemophilus parainfluenzae*.

Concentration: 5000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No characteristic banding pattern change after 16h incubation of 1 µg of λ DNA with 10 units of enzyme at 37°C.



HspA I

| HspA I | Cat# | Pack Size |
|--------|--------|-----------|
| G↓CGC | 279010 | 1000 U |
| CGC↑G | 279050 | 5000 U |

Purified from *Haemophilus species A1*.

Isoschizomer: Hin6 I.

Concentration: 10000-30000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 30 units of enzyme at 37°C.



Kpn I

| Kpn I | Cat# | Pack Size |
|---------|--------|-----------|
| GGTAC↓C | 227020 | 2000 U |
| C↑CATGG | 227100 | 10000 U |

Purified from *Klebsiella pneumoniae*.

Concentration: 20000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 70% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 30 units of enzyme at 37°C.



Restriction Endonucleases

| Ksp22 I | Cat# | Pack Size |
|---------|--------|-----------|
| T↓GATCA | 228010 | 1000 U |
| ACTAG↑T | 228050 | 5000 U |

Ksp22 I

Purified from *Kurthia species 22*.

Isoschizomer: Bcl I.

Concentration: 20000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 70% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of the enzyme at 37°C.



| Kzo9 I | Cat# | Pack Size |
|--------|--------|-----------|
| ↓GATC | 229002 | 200 U |
| CTAG↑ | 229010 | 1000 U |

Kzo9 I

Purified from *Kurthia zopfii 9*.

Isoschizomer: Sau3A I.

Concentration: 2000-5000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 95% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 4 units of the enzyme at 37°C.



| Mlu I | Cat# | Pack Size |
|---------|--------|-----------|
| A↓CGCGT | 230010 | 1000 U |
| TGCGC↑A | 230050 | 5000 U |

Mlu I

Purified from *Micrococcus luteus*.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of enzyme at 37°C.



| MroN I | Cat# | Pack Size |
|---------|--------|-----------|
| G↓CCGGC | 231005 | 500 U |
| CGGCC↑G | 231025 | 2500 U |

MroN I

Purified from *Micrococcus roseus NO*.

Concentration: 5000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments of pBR322 may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of Adenovirus-2 DNA with 4 units of enzyme at 37°C.



| | | | | | | | | | | | |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|
| 37°C | Optimal reaction temperature | λ | Substrate DNA for activity test | O | Optimal reaction buffer | Yes | Inactivation at 65°C, 20 min | No | No Inactivation at 65°C, 20 min | 80°C | Inactivation at 80°C, 20 min |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|

Msp I

| Msp I | Cat# | Pack Size |
|-------|--------|-----------|
| C↓CGG | 232010 | 1000 U |
| GGC↑C | 232050 | 5000 U |

Purified from *Moraxella species*.

Concentration: 10000-30000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of the enzyme at 37°C.



Nru I

| Nru I | Cat# | Pack Size |
|---------|--------|-----------|
| TCC↓CGA | 233005 | 500 U |
| AGC↑GCT | 233025 | 2500 U |

Purified from *Nocardia rubra*.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 3-fold overdigestion with the enzyme, more than 10% of the DNA fragments may be ligated.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 15 units of the enzyme at 37°C.



Pce I

| Pce I | Cat# | Pack Size |
|---------|--------|-----------|
| AGG↓CCT | 236010 | 1000 U |
| TCC↑GGA | 236050 | 5000 U |

Purified from *Planococcus citreus 55*.

Isoschizomer: Stu I.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 70% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of the enzyme at 50°C.



Pci I

| Pci I | Cat# | Pack Size |
|---------|--------|-----------|
| A↓CATGT | 234002 | 200 U |
| TGTAC↑A | 234010 | 1000 U |

Purified from *Planococcus citreus SE-F45*.

Isoschizomer: BspLU11 I.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of T7 DNA with 20 units of the enzyme at 37°C.



Restriction Endonucleases

| Ple19 I | Cat# | Pack Size |
|---------|--------|-----------|
| CGAT↓CG | 235001 | 100 U |
| GC↑TAGC | 235005 | 500 U |

Ple19 I

Purified from *Pseudomonas lemoignei* 19.

Isoschizomer: Pvu I.

Concentration: 5000 units/ml.

Ligation/recutting assay:

After 2-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 10 units of the enzyme at 37°C.



| PspOM I | Cat# | Pack Size |
|---------|--------|-----------|
| G↓GGCC | 280005 | 1500 U |
| CCCGG↑C | 280025 | 7500 U |

PspOM I

Purified from *Pseudomonas species OM*.

Concentration: 20000-50000 units/ml.

Ligation/recutting assay:

After 40-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 100 units of enzyme at 37°C.



| Psp124B I | Cat# | Pack Size |
|-----------|--------|-----------|
| GAGCT↓C | 237010 | 1000 U |
| C↑TCGAG | 237050 | 5000 U |

Psp124B I

Purified from *Pseudomonas species 124B*.

Isoschizomer: Sac I.

Concentration: 20000-50000 units/ml.

Ligation/recutting assay:

After 40-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 40 units of the enzyme at 37°C.



| PspE I | Cat# | Pack Size |
|----------|--------|-----------|
| G↓GTNACC | 238020 | 2000 U |
| CCANTG↑G | 238100 | 10000 U |

PspE I

Purified from *Pseudomonas species E*.

Isoschizomer: BstE II.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of the enzyme at 37°C.



| | | | | | | | | | | | |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|
| 37°C | Optimal reaction temperature | λ | Substrate DNA for activity test | O | Optimal reaction buffer | Yes | Inactivation at 65°C, 20 min | No | No Inactivation at 65°C, 20 min | 80°C | Inactivation at 80°C, 20 min |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|



Pst I

| Pst I | Cat# | Pack Size |
|---------|--------|-----------|
| CTGCA↓G | 239040 | 4000 U |
| G↑ACGTC | 239200 | 20000 U |

Purified from *E.coli* strain that carries Pst I gene from *Providencia stuartii*.

Concentration: 20000-50000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 60 units of the enzyme at 37°C.



Pvu II

| Pvu II | Cat# | Pack Size |
|---------|--------|-----------|
| CAG↓CTG | 240020 | 2000 U |
| GTC↑GAC | 240100 | 10000 U |

Purified from *Proteus vulgaris*.

Concentration: 20000-50000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 70% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 40 units of the enzyme at 37°C.



Rsa I

| Rsa I | Cat# | Pack Size |
|-------|--------|-----------|
| GT↓AC | 241010 | 1000 U |
| CA↑TG | 241050 | 5000 U |

Purified from *Rhodospseudomonas sphaeroides*.

Concentration: 10000-30000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 40 units of the enzyme at 37°C.



Sal I

| Sal I | Cat# | Pack Size |
|---------|--------|-----------|
| G↓TCGAC | 242020 | 2000 U |
| CAGCT↑G | 242100 | 10000 U |

Purified from *Streptomyces albus*.

Concentration: 20000 units/ml.

Ligation/recutting assay:

After 20-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 60 units of the enzyme at 37°C.



Restriction Endonucleases

Sbf I

| Sbf I | Cat# | Pack Size |
|-----------|--------|-----------|
| CCTGCA↓GG | 243002 | 200 U |
| GG↑ACGTCC | 243010 | 1000 U |

Purified from *Streptomyces species Bf6I*.

Isoschizomer: Sse8387 I.

Concentration: 5000 units/ml.



Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 10 units of the enzyme at 37°C.

Sfi I

| Sfi I | Cat# | Pack Size |
|----------------|--------|-----------|
| GGCCNNNN↓NGGCC | 244005 | 500 U |
| CCGGN↑NNNNCCGG | 244025 | 2500 U |

Purified from *Streptomyces fimbriatus*.

Concentration: 2000-15000 units/ml.



Ligation/recutting assay:

After 3-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of Adenovirus-2 DNA with 2 units of the enzyme at 37°C.

Sfr274 I

| Sfr274 I | Cat# | Pack Size |
|----------|--------|-----------|
| C↓TCGAG | 245020 | 2000 U |
| GAGCT↑C | 245100 | 10000 U |

Purified from *Streptomyces fradiae 274*.

Isoschizomer: Xho I.

Concentration: 50000 units/ml.



Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 5 units of the enzyme at 37°C.

Sfr303 I

| Sfr303 I | Cat# | Pack Size |
|----------|--------|-----------|
| CCGC↓GG | 281010 | 1000 U |
| GG↑CGCC | 281050 | 5000 U |

Purified from *Streptomyces fradiae 303*.

Isoschizomer: Sac II.

Concentration: 10000 units/ml.



Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 10 units of enzyme at 37°C.

| | | | | | | | | | | | |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|
| 37°C | Optimal reaction temperature | λ | Substrate DNA for activity test | B | Optimal reaction buffer | Yes | Inactivation at 65°C, 20 min | No | No Inactivation at 65°C, 20 min | 80°C | Inactivation at 80°C, 20 min |
|------|------------------------------|---|---------------------------------|---|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|

Sma I

| Sma I | Cat# | Pack Size |
|---------|--------|-----------|
| CCC↓GGG | 246020 | 2000 U |
| GGG↑CCC | 246100 | 10000 U |

Purified from *E.coli* strain that carries SmaI gene from *Serratia marcescens*.

Concentration: 20000 units/ml.

Ligation/recutting assay:

After 2-fold overdigestion with the enzyme, more than 30% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 25 units of the enzyme at 25°C.



Smi I

| Smi I | Cat# | Pack Size |
|-----------|--------|-----------|
| ATTT↓AAAT | 247010 | 1000 U |
| TAAA↑TTTA | 247050 | 5000 U |

Purified from *Streptococcus milleri* S.

Isoschizomer: Swa I.

Concentration: 20000-50000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 80% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of T7 DNA with 40 units of the enzyme at 25°C.



Sph I

| Sph I | Cat# | Pack Size |
|----------|--------|-----------|
| GCA TG↓C | 248002 | 200 U |
| C↑GTACG | 248010 | 1000 U |

Purified from *Sphaerotilus species*.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 10 units of the enzyme at 37°C.



Sse9 I

| Sse9 I | Cat# | Pack Size |
|--------|--------|-----------|
| ↓AATT | 249001 | 100 U |
| TTAA↑ | 249005 | 500 U |

Purified from *Sporosarcina species* 9.

Concentration: 1000-5000 units/ml.

Ligation/recutting assay:

After 2-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 8 units of the enzyme at 55°C.



| | | | | | | | |
|------------------------------|---------------------------------|-------------------------|-----|---------------------------------|-----|---------------------------------|-----------------------------------|
| Optimal reaction temperature | Substrate DNA for activity test | Optimal reaction buffer | Yes | No Inactivation at 65°C, 20 min | Yes | No Inactivation at 65°C, 20 min | 80°C Inactivation at 80°C, 20 min |
|------------------------------|---------------------------------|-------------------------|-----|---------------------------------|-----|---------------------------------|-----------------------------------|

Restriction Endonucleases

| Ssp I | Cat# | Pack Size |
|---------|--------|-----------|
| AAT↓ATT | 250005 | 500 U |
| TTA↑TAA | 250025 | 2500 U |

Purified from *Sphaerotilus species*.

Concentration: 5000 units/ml.

Ssp I

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 5 units of the enzyme at 37°C.



| Taq I | Cat# | Pack Size |
|-------|--------|-----------|
| T↓CGA | 251002 | 200 U |
| AGC↑T | 252010 | 1000 U |

Purified from *Thermus aquaticus*.

Concentration: 5000 units/ml.

Taq I

Ligation/recutting assay:

After 3-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 6 units of the enzyme at 65°C.



| Tru9 I | Cat# | Pack Size |
|--------|--------|-----------|
| T↓TAA | 252002 | 200 U |
| AAT↑T | 252010 | 1000 U |

Purified from *Thermus ruber 9*.

Concentration: 20000-50000 units/ml.

Tru9 I

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 95% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 40 units of the enzyme at 65°C.



| Vha464 I | Cat# | Pack Size |
|----------|--------|-----------|
| C↓TTAAG | 282005 | 500 U |
| GAATT↑C | 282025 | 2500 U |

Purified from *Vibrio harveyi 464*.

Concentration: 20000 units/ml.

Vha464 I

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 40% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of enzyme at 37°C.



| | | | | | | | | | | | |
|------|------------------------------|---|---------------------------------|------------------|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|
| 37°C | Optimal reaction temperature | λ | Substrate DNA for activity test | Erlenmeyer flask | Optimal reaction buffer | Yes | Inactivation at 65°C, 20 min | No | No Inactivation at 65°C, 20 min | 80°C | Inactivation at 80°C, 20 min |
|------|------------------------------|---|---------------------------------|------------------|-------------------------|-----|------------------------------|----|---------------------------------|------|------------------------------|



Vne I

| Vne I | Cat# | Pack Size |
|---------|--------|-----------|
| G↓TGCAC | 253010 | 1000 U |
| CACGT↑G | 253050 | 5000 U |

Purified from *Vibrio nereis* 18.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of the enzyme at 37°C.



Vsp I

| Vsp I | Cat# | Pack Size |
|---------|--------|-----------|
| AT↓TAAT | 254010 | 1000 U |
| TAAT↑TA | 254050 | 5000 U |

Purified from *Vibrio species* 343.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 70% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 10 units of the enzyme at 37°C.



Xba I

| Xba I | Cat# | Pack Size |
|---------|--------|-----------|
| T↓CTAGA | 255010 | 1000 U |
| AGATC↑T | 255050 | 5000 U |

Purified from *Xanthomonas badrii*.

Concentration: 20000-50000 units/ml.

Ligation/recutting assay:

After 10-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of

1 µg of λ DNA with 20 units of the enzyme at 37°C.



Zsp2 I

| Zsp2 I | Cat# | Pack Size |
|---------|--------|-----------|
| ATGCA↓T | 256010 | 1000 U |
| T↑ACGTA | 256050 | 5000 U |

Purified from *Zoogloea species* 2.

Isoschizomer: Nsi I.

Concentration: 10000 units/ml.

Ligation/recutting assay:

After 5-fold overdigestion with the enzyme, more than 90% of the DNA fragments may be ligated and recut.

Nonspecific endonuclease test:

No change of characteristic banding pattern after 16h incubation of 1 µg of λ DNA with 20 units of the enzyme at 37°C.



Restriction Endonucleases, Activity Table

| Enzyme | Recog. Sequence | SEBuffer | BSA | Activity (% from maximum) | | | | | t,0' 37°C | Inactivation, 20 min |
|-----------|-----------------------------|----------|-----|---------------------------|--------|--------|--------|--------|--------------|-------------------------|
| | | | | B | G | O | W | Y | | |
| Aau I | T [^] GTACA | W | + | 50-75 | 75-100 | 50-75 | 100 | 75-100 | 37 | 65°C |
| Acc113 I | AGT [^] ACT | Y | + | 50-75 | 25-50 | 0-10 | 0-10 | 100 | 37 | 65°C |
| Acc16 I | TGC [^] GCA | W | - | 50-75 | 75-100 | 25-50 | 100 | 75-100 | 37 | 65°C |
| Acc65 I | G [^] GTACC | W | - | 10-25 | 25-50 | 75-100 | 100 | 10-25 | 37 | 65°C |
| AccBS I | GAG [^] CGG | Y | - | 75-100 | 75-100 | 25-50 | 25-50 | 100 | 37 | 65°C |
| Acs I | R [^] AAATTY | W | + | 25-50 | 50-75 | 50-75 | 100 | 10-25 | 50 | 80°C |
| Afe I | AGC [^] GCT | Y | - | 10-25 | 25-50 | 75-100 | 75-100 | 100 | 37 | 65°C |
| Ahl I | A [^] CTAGT | B | + | 100 | 75-100 | 25-50 | 25-50 | 75-100 | 37 | 80°C |
| Alu I | AG [^] CT | Y | - | 75-100 | 75-100 | 10-25 | 50-75 | 100 | 37 | 65°C |
| Apa I | GGGCC [^] C | Y | + | 50-75 | 25-50 | 0-10 | 0-10 | 100 | 37 | 65°C |
| AspS9 I | G [^] GNCC | W | - | 50-75 | 50-75 | 75-100 | 100 | 50-75 | 37 | 65°C |
| AsuNH I | G [^] CTAGC | Y | + | 75-100 | 50-75 | 0-10 | 0-10 | 100 | 37 | 65°C |
| BamH I | G [^] GATCC | G | + | 25-50 | 100 | 75-100 | 75-100 | 25-50 | 37 | 65°C |
| Bgl I | GCCNNNN [^] NGGC | 2W | - | 50-75 | 50-75 | 0-10 | 75-100 | 25-50 | 37 | 65°C |
| Bgl II | A [^] GATCT | O | - | 0-10 | 10-25 | 100 | 25-50 | 10-25 | 37 | 80°C |
| Bme18 I | G [^] GWCC | O | - | 10-25 | 25-50 | 100 | 75-100 | 10-25 | 37 | 65°C |
| Bpu14 I | TT [^] CGAA | G | - | 50-75 | 100 | 25-50 | 25-50 | 75-100 | 37 | 65°C |
| Bsa29 I | AT [^] CGAT | G | + | 25-50 | 100 | 50-75 | 50-75 | 75-100 | 37 | 65°C |
| Bsc4 I | CCNNNN [^] NNGG | W | + | 75-100 | 75-100 | 50-75 | 100 | 25-50 | 55 | 80°C |
| BseI I | ACTGG(1/-1) | Y | - | 75-100 | 75-100 | 25-50 | 10-25 | 100 | 65 | 80°C |
| Bse118 I | R [^] CCGGY | O | - | 0-10 | 50-75 | 100 | 75-100 | 25-50 | 65 | 80°C |
| Bse21 I | CC [^] TNAGG | Y | - | 50-75 | 50-75 | 10-25 | 25-50 | 100 | 37 | 80°C |
| Bse8 I | GATNN [^] NNATC | G | - | 25-50 | 100 | 75-100 | 75-100 | 50-75 | 60 | 80°C |
| Bsp19 I | C [^] CATGG2 | W | + | 0-10 | 10-25 | 50-75 | 75-100 | 10-25 | 37 | 65°C |
| BssNA I | GTA [^] TAC | W | + | 50-75 | 50-75 | 75-100 | 100 | 75-100 | 37 | 80°C |
| BssT1 I | C [^] CWWGG | 2K | - | 10-25 | 25-50 | 25-50 | 75-100 | 10-25 | 60 | 80°C |
| Bst4C I | ACN [^] GT | Y | - | 75-100 | 75-100 | 10-25 | 25-50 | 100 | 65 | 80°C |
| BstDS I | C [^] CRYGG | Y | - | 0-10 | 75-100 | 50-75 | 25-50 | 100 | 65 | 80°C |
| BstFN I | CG [^] CG | Y | - | 75-100 | 50-75 | 25-50 | 25-50 | 100 | 60 | 80°C |
| BstH2 I | RGCGC [^] Y | Y | + | 50-75 | 50-75 | 0-10 | 10-25 | 100 | 65 | 80°C |
| BstHH I | GCG [^] C | Y | + | 75-100 | 50-75 | 25-50 | 50-75 | 100 | 50 | 65°C |
| BstHP I | GTT [^] AAAC | Y | + | 0-10 | 50-75 | 10-25 | 25-50 | 100 | 37 | 65°C |
| BsuR I | GG [^] CC | G | - | 75-100 | 100 | 25-50 | 50-75 | 50-75 | 37 | 80°C |
| CciN I | GC [^] GGCCGC | Y | - | 25-50 | 50-75 | 75-100 | 75-100 | 100 | 37 | 65°C |
| Dra I | TTT [^] AAA | G | + | 75-100 | 100 | 25-50 | 75-100 | 75-100 | 37 | 65°C |
| EcoR I | G [^] AATTC | * | + | 50-75 | 75-100 | 75-100 | 100 | 50-75 | 37 | 65°C |
| EcoR V | GAT [^] ATC | W | + | 0-10 | 25-50 | 50-75 | 100 | 25-50 | 37 | 80°C |
| FauND I | CA [^] TATG | Y | + | 50-75 | 75-100 | 10-25 | 50-75 | 100 | 37 | 65°C |
| Hind III | A [^] AGCTT | W | + | 10-25 | 25-50 | 0-10 | 100 | 0-10 | 37 | 80°C |
| Hinf I | G [^] ANTC | O | - | 25-50 | 75-100 | 100 | 75-100 | 75-100 | 37 | 80°C |
| Hpa I | GTT [^] AAC | Y | - | 0-10 | 50-75 | 10-25 | 25-50 | 100 | 37 | 65°C |
| HspA I | G [^] CGC | Y | - | 50-75 | 50-75 | 25-50 | 25-50 | 100 | 37 | 80°C |
| Kpn I | GGTAC [^] C | B | + | 100 | 25-50 | 25-50 | 25-50 | 75-100 | 37 | 80°C |
| Ksp22 I | T [^] GATCA | G | + | 50-75 | 100 | 50-75 | 50-75 | 25-50 | 37 | 65°C |
| Kzo9 I | [^] GATC | G | + | 50-75 | 100 | 50-75 | 50-75 | 50-75 | 37 | 65°C |
| Mlu I | A [^] CGCGT | O | - | 0-10 | 10-25 | 100 | 25-50 | 10-25 | 37 | 65°C |
| MroN I | G [^] CCGGC | B | - | 100 | 50-75 | 10-25 | 0-10 | 10-25 | 37 | 80°C |
| Msp I | C [^] CGG | B | - | 100 | 75-100 | 50-75 | 75-100 | 75-100 | 37 | 65°C |
| Nru I | TCG [^] CGA | W | - | 0-10 | 10-25 | 75-100 | 100 | 10-25 | 37 | 80°C |
| Pee I | AGG [^] CCT | Y | - | 75-100 | 75-100 | 50-75 | 25-50 | 100 | 50 | 80°C |
| Pci I | A [^] CATGT | O | - | 50-75 | 75-100 | 100 | 75-100 | 50-75 | 37 | 65°C |
| Ple19 I | CGAT [^] CG | Y | - | 75-100 | 75-100 | 25-50 | 25-50 | 100 | 37 | 65°C |
| Psp124B I | GAGCT [^] C | G | - | 75-100 | 100 | 10-25 | 0-10 | 75-100 | 37 | 80°C |
| PspE I | G [^] GTNACC | B | - | 100 | 50-75 | 25-50 | 50-75 | 50-75 | 37 | 65°C |
| PspOM I | G [^] GGCCC | Y | - | 75-100 | 10-25 | 0-10 | 0-10 | 100 | 37 | 65°C |
| Pst I | CTGCA [^] G | O | + | 10-25 | 25-50 | 100 | 25-50 | 25-50 | 37 | 80°C |
| Pvu II | CAG [^] CTG | G | + | 25-50 | 100 | 25-50 | 25-50 | 25-50 | 37 | 80°C |
| Rsa I | GT [^] AC | B | - | 100 | 50-75 | 0-10 | 50-75 | 75-100 | 37 | 80°C |
| Sal I | G [^] TCGAC | O | - | 0-10 | 10-25 | 100 | 25-50 | 0-10 | 37 | 65°C |
| Sbf I | CCTGCA [^] GG | Y | - | 75-100 | 50-75 | 0-10 | 0-10 | 100 | 37 | 80°C |
| Sfi I | GGCCNNNN [^] NGGCC | G | + | 75-100 | 100 | 25-50 | 25-50 | 25-50 | 50 | 65°C |
| Sfr274 I | C [^] TCGAG | B | - | 100 | 75-100 | 50-75 | 50-75 | 75-100 | 50 | 65°C |
| Sfr303 I | CCGC [^] GG | B | - | 100 | 50-75 | 10-25 | 10-25 | 75-100 | 37 | 65°C |
| Sma I | CCC [^] GGG | Y | - | 0-10 | 0-10 | 0-10 | 0-10 | 100 | 25 | 65°C |
| Smi I | ATTT [^] AAAT | O | + | 25-50 | 25-50 | 100 | 75-100 | 25-50 | 37 | 65°C |
| Sph I | GCATG [^] C | G | + | 25-50 | 100 | 75-100 | 75-100 | 50-75 | 37 | 65°C |
| Sse9 I | [^] AATT | B | + | 100 | 75-100 | 50-75 | 50-75 | 75-100 | 55 | 65°C |
| Ssp I | AAT [^] ATT | K | - | 75-100 | 50-75 | 25-50 | 50-75 | 75-100 | 37 | 65°C |
| Taq I | T [^] CGA | Y | + | 50-75 | 75-100 | 75-100 | 50-75 | 100 | 65 | 80°C |
| Tru9 I | T [^] TAA | W | - | 75-100 | 25-50 | 25-50 | 100 | 50-75 | 65 | 80°C |
| Vha464 I | C [^] TAAAG | G | - | 50-75 | 100 | 25-50 | 10-25 | 75-100 | 37 | 65°C |
| Vne I | G [^] TGCAC | O | - | 10-25 | 25-50 | 100 | 25-50 | 25-50 | 37 | 65°C |
| Vsp I | AT [^] TAAT | W | - | 0-10 | 10-25 | 50-75 | 100 | 25-50 | 37 | 65°C |
| Xba I | T [^] CTAGA | O | - | 75-100 | 75-100 | 100 | 50-75 | 75-100 | 37 | 65°C |
| Zsp2 I | ATGCA [^] T | B | + | 100 | 50-75 | 25-50 | 25-50 | 25-50 | 37 | 65°C |
| N.Bst9 I | GAGTC(4/-) | * | - | 10-25 | 75-100 | 100 | 100 | 50-75 | 55 | 80°C |